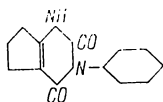


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MICROBIOLOGICAL TRANSFORMATION OF HERBICIDES
IN CATABOLIC PROCESSES

PART I. THE EFFECT OF VENZAR ON THE RESPIRATION
ACTIVITY OF SELECTED GENERA OF SOIL BACTERIA ¹

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Venzar (3-cyclohexyl-5, 6-trimethyleneuracil) belongs to

herbicides slightly soluble in water (at 25° 6 ppm). To weeds it is a photosynthesis inhibitor. Soil bacteria are little susceptible to this herbicide.

In the presence of *Bacillus* sp. venzar markedly increased the respiration and decreased the photosynthesis of leaves of *Sinapis alba* and *Spirodella polyrrhiza* as compared with the action of venzar itself and control [8]. Our aim was to test whether and to what extent venzar can influence the metabolism of bacteria resistant to high doses of this herbicide, and particularly their respiration activity.

MATERIAL AND METHODS

The tested bacterial strains were isolated from previously percolated soils: sandy-loamy soil, the Wrocław black earth, sandy soil Yugoslavian black-earth treated with low doses of venzar (1.5 ppm). The origin of individual bacterial strains, their genus affiliation as well as the conditions of isolation are given in Table 1. To isolate the strains of bacteria under examination the following media were chosen: the Kaufman synthetic medium [12], the Kearney synthetic medium [11], the Čapek medium [7] for *Pseudomonas*, Sands and Rovira [15] and a soil extract (flood with 1.5 l of tap water 1 kg of soil, sterilize after

¹ Supported financially by the USA Department of Agriculture. Res. Service Grant Fg. Po — 335. Presented at International Symposium "Testing of chemical substances for exotoxicological evaluation," May 1979 GSP, Munich—Neuherberg.

Table 1

List and origin of bacterial strains tested

Bacterial strain		Percolated soil + venzar	ppm	Culture medium for isolation + venzar	ppm
Symbol	Genus				
V-217	<i>Bacillus</i> sp.	loamy-sandy	0	Kearney	1000
V-218	<i>Bacillus</i> sp.	loamy-sandy+2% cellulose	0	Kearney	0
V- 92	<i>Bacillus</i> sp.	black degraded	1.5	Kearney	100
V- 25	<i>Bacillus</i> sp.	loamy-sandy	0	soil extract	100
V- 76	<i>Bacillus</i> sp.	loamy-sandy fertilized	3	Capek	100
V- 20	<i>Bacillus</i> sp.	loamy-sandy	9	synthetic, Kaufman	100
V- 16	<i>Bacillus mycoides</i>	loamy-sandy fertilized	3	Kaufman synthetic	100

V- 80	<i>Pseudomonas fluorescens</i>	Yugoslav black-earth	1.5	for <i>Pseudomonas</i>	1000
V-209	<i>Pseudomonas fluorescens</i>	Yugoslav black-earth+2% cellulose	1.5	for <i>Pseudomonas</i>	100
V-208	<i>Pseudomonas fluorescens</i>	Yugoslav black-earth+2% cellulose	1.5	for <i>Pseudomonas</i>	1000
V-105	<i>Pseudomonas fluorescens</i>	black degraded	1.5	Kearney	100
V- 65	<i>Pseudomonas fluorescens</i>	loamy-sandy fertilized	9.0	for <i>Pseudomonas</i>	100
V-212	<i>Pseudomonas aeruginosa</i>	Yugoslav black-earth+2% cellulose	0.0	for <i>Pseudomonas</i>	1000
V-204	<i>Pseudomonas aeruginosa</i>	loamy-sandy	1.5	for <i>Pseudomonas</i>	100
V-211	<i>Pseudomonas aeruginosa</i>	sandy	1.5	for <i>Pseudomonas</i>	1000
V-100	<i>Pseudomonas aeruginosa</i>	black degraded	1.5	for <i>Pseudomonas</i>	1000
VI-5	<i>Pseudomonas aeruginosa</i>	loamy-sandy	0.0	for <i>Pseudomonas</i>	100
V-206	<i>Pseudomonas aeruginosa</i>	Yugoslav black-earth	1.5	for <i>Pseudomonas</i>	1000
V-223	<i>Pseudomonas aeruginosa</i>	loamy-sandy	1.5	for <i>Pseudomonas</i>	1000
V-23a	<i>Pseudomonas aeruginosa</i>	loamy-sandy fertilized	1.5	Kearney	1000
V-205	<i>Pseudomonas aeruginosa</i>	loamy-sandy	1.5	for <i>Pseudomonas</i>	0
V-11	<i>Pseudomonas alcaligenes</i>	loamy-sandy	9.0	Kaufman synthetic	1000
V-1	<i>Pseudomonas alcaligenes</i>	loamy-sandy	9.0	Kaufman synthetic	0
VIV-3	<i>Pseudomonas species</i>	black degraded fertilized	0.0	Kearney	1000

V-4a	<i>Corynebacterium</i> sp.	loamy-sandy	9.0	Kaufman synthetic	1000
V-84	<i>Corynebacterium</i> sp.	sandy	1.5	Kearney	100
V-36	<i>Corynebacterium</i> sp.	loamy-sandy	0	soil extract	100

V-216	<i>Arthrobacter</i> sp.	loamy-sandy	0	Kearney	1000
VVI-8	<i>Arthrobacter</i> sp.	black degraded fertilized	0	Kearney	100

V-5	<i>Micrococcus</i> sp.	loamy-sandy	9.0	Kaufman synthetic	100

VIX-1	<i>Serratia</i> sp.	black degraded fertilized	0	Kearney	1000

V-10	<i>Nocardia</i> sp.	loamy-sandy fertilized	0	for <i>Pseudomonas</i>	100

24 hours and filter the liquid through filter paper and enrich with KH_2PO_4 — 0.5 g/l, glucose — 10 g and agar 16 g, and then sterilize again). A part of it was enriched with 100 and 1000 ppm of venzar. A corresponding herbicide dose was suspended in a small volume ethanol and added to previously sterilized media. 32 bacterial strain coming from four successive microbiological analyses were examined, that is: 17 strains of genus *Pseudomonas*, 7 strains of genus *Bacillus*, 5 strains of genus *Corynebacterium*, one *Nocardia*, sp., one *Serratia* sp. and one *Micrococcus* sp.

The respiration measurements were made using the respirometric

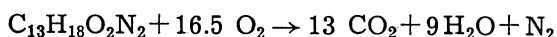
method in the Warburg apparatus at 27°C in vessels of 3 ml volume [5, 6]. One-day liquid broth cultures of individual bacterial strains were centrifuged (10 000 r.p.), and washed with physiological salt solution, as well as suspended in a phosphate buffer of pH 7.2. One millilitre of this suspension was introduced into the main chamber of the vessel. The same amount of suspension was dried at 105°C up to a constant weight in order to determine the dry mass of cells which was between 10 and 12 mg/ml. Into the side arms of the Warburg vessels the respiration substrates were introduced: a solution of watersoluble dose of venzar (6 ppm) and 0.1% glucose solution, or mixtures of both these compounds. For each bacterial strain the oxygen uptake and carbon dioxide evolution in the presence of the above-mentioned respiration substrates were examined, and the respiration factors RQ were calculated. The measurements were usually carried out over 8 hours, reading out the level of manometers every half an hour [17]. Apart from the exogenic respiration, endogenic respiration was also examined. The results of the gas exchange were expressed in microlitres (μl) of the taken up or evolved gas for one mg of dry mass of bacteria. On the basis of the obtained results the oxygen uptake curves and the carbon dioxide evolution curves were plotted.

By the method of thin-layer chromatography the undegraded herbicide and its metabolites in the liquid from the Warburg vessels was determined after completion of gas exchange investigation. Venzar was extracted with ethyl acetate, and the chromatographic plates covered with silica gel G-60 were developed in the system benzene:acetone (6:1) and then sprayed with a starch reagent [14].

RESULTS AND DISCUSSION

The effect of venzar on the respiration activity of various genera of bacteria was different (Figs 1-7). Most bacteria of genera *Bacillus* (Fig. 2), *Pseudomonas* and *Arthrobacter* (Figs 1, 4) show ability to oxidation of venzar to equal degree as glucose and in some cases venzar was even better utilized (Figs 1, 2). The bacteria of genus *Pseudomonas aeruginosa* and one strain *Arthrobacter* sp. very intensely oxidated the mixtures of glucose and venzar (Fig. 4).

The respiration factors RQ for bacteria oxidating venzar were higher than this would result from the stochiometric reaction (Table 2)



$$\text{RQ} = \frac{13 \text{ CO}_2}{16.5 \text{ O}_2} = 0.78$$

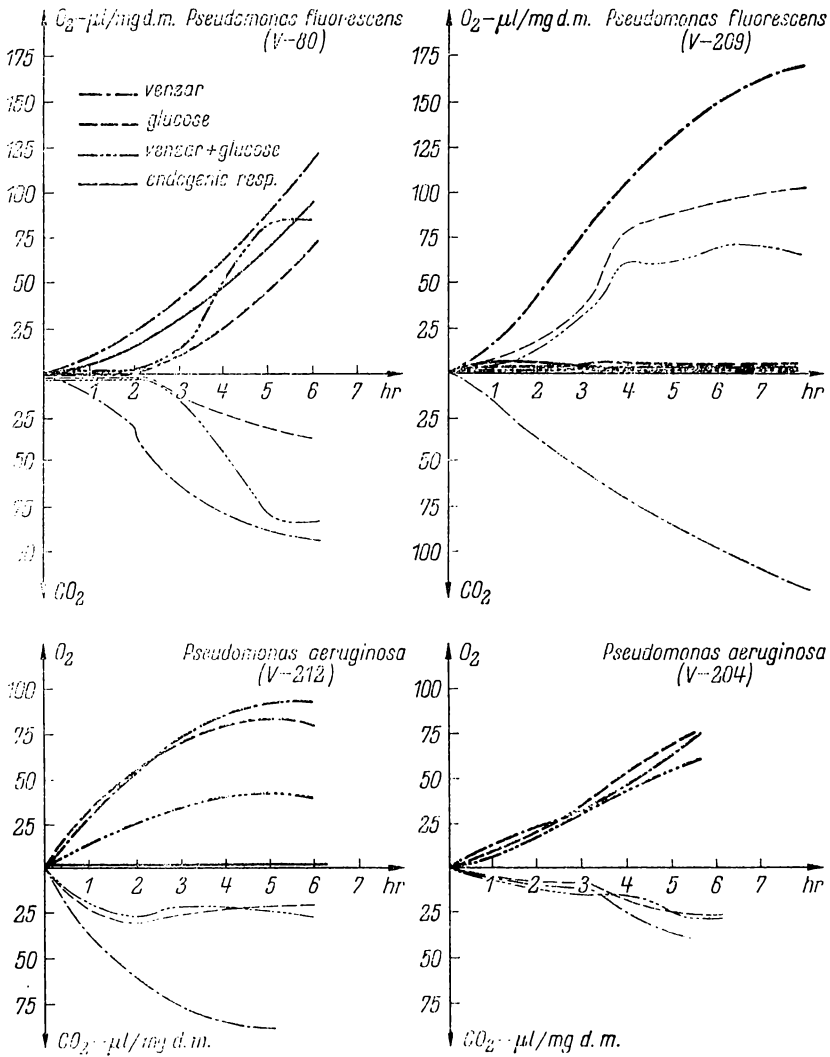


Fig. 1. The respiration of *Pseudomosa* strains utilizing various carbon sources

The genus *Pseudomonas* prevailed quantitatively in the population of isolated bacteria resistant to high doses of venzar in the substrate (100 and 1000 ppm) and most intensely utilized this herbicide as a carbon source (Fig. 1). After completion of the investigation of gas exchange in the liquid from the Warburg vessels, in most cases no residues of undegraded herbicide were detected, while there appeared new metabolites (Table 2). For the genus *Bacillus* venzar was also a carbon source. A decrement of about 75% of this herbicide was noted chromatographically but no metabolites were detected. Factors RQ were higher for venzar, which could indicate the predominance of decarbo-

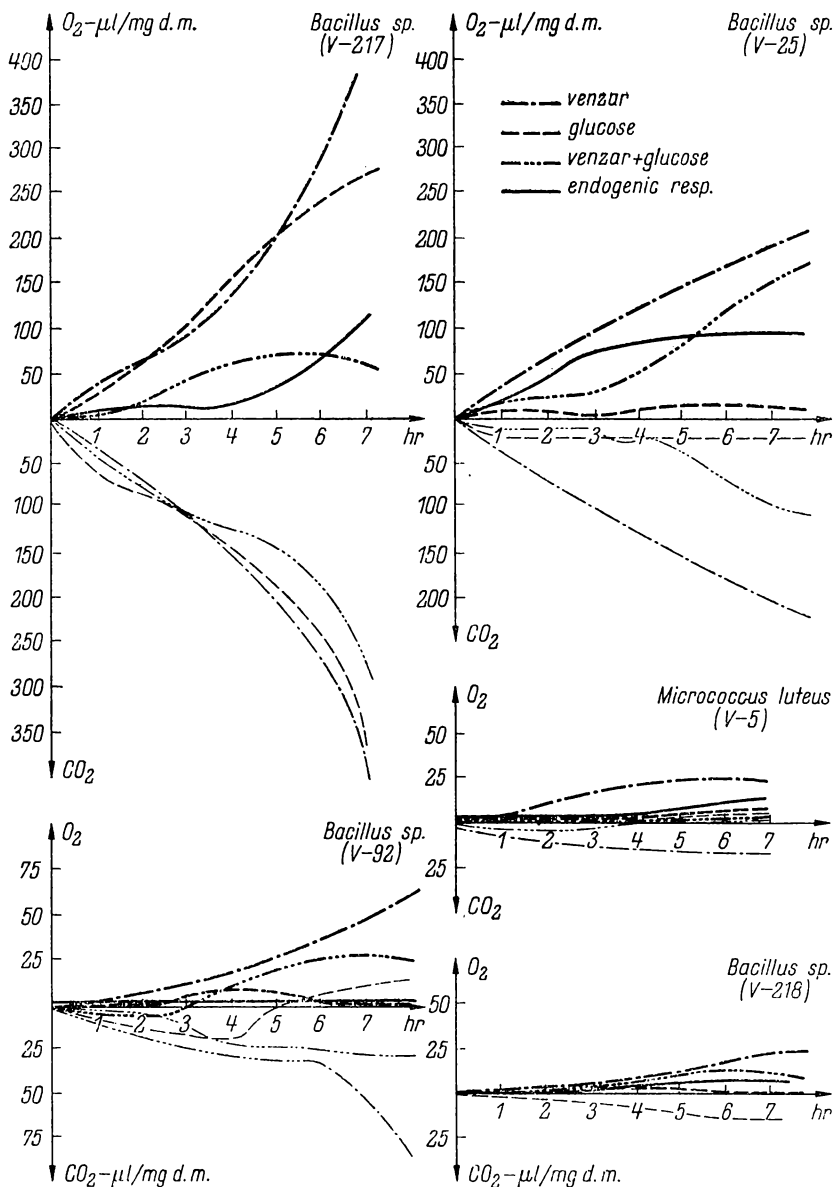


Fig. 2. The respiration of *Bacillus* sp. and *Micrococcus* sp. strains—various carbon sources

xylation processes, incomplete oxidation of the herbicide and of the evolution of yet other gases besides CO_2 , or of oxidation typical of organic acids which could be formed in the degradation process of venzar. It is not known which of the possibilities mentioned was the cause of increased values of the respiration factors RQ.

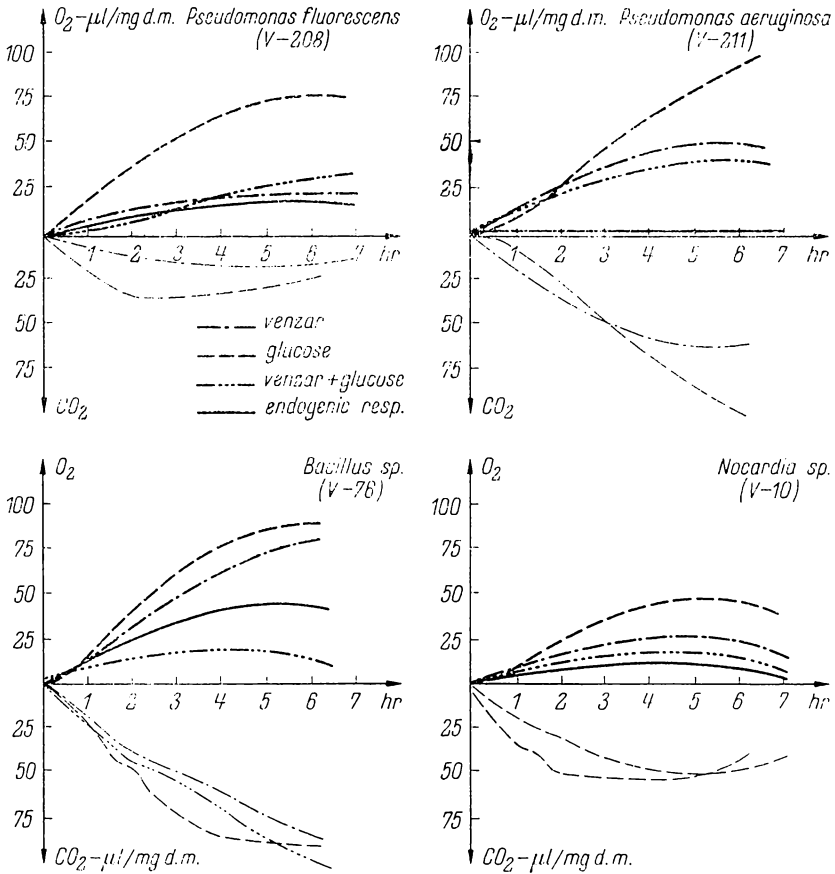


Fig. 3. The effect of glucose and venzar on the respiration of various bacterial strains

The other genera of bacteria: *Corynebacterium*, *Serratia*, *Nocardia*, *Micrococcus*, *Arthrobacter* and *Pseudomonas alcaligenes* oxidated venzar more poorly (Figs. 2, 5, 6, 7) or they could not adapt themselves to the utilization of venzar as a respiration substrate. Thus they have shown a very small respiration activity. At the same time a disturbance in respiration processes was observed, manifesting itself by breakdown and stoppage of respiration, the respiration factors RQ reaching in these experiments irrational values (Table 2). The above-mentioned results indicate that venzar changed the metabolism of these bacterial strains

A water-soluble dose of venzar (6 ppm) acted stimulating on the respiration of some of bacteria of genera *Pseudomonas*, *Bacillus* and *Arthrobacter* (Figs. 1, 2, 4), the stimulation being proportional to both the oxygen uptake and CO_2 evolution. Other herbicides, as reported

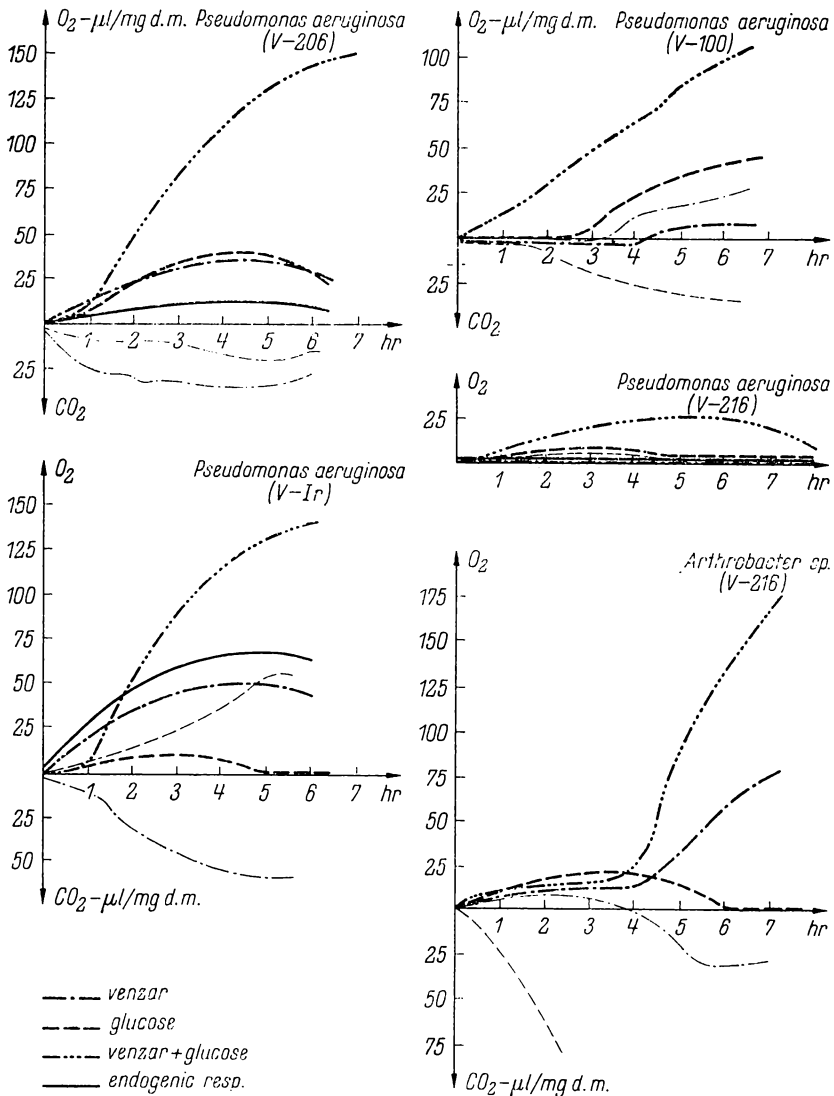


Fig. 4. The effect of glucose and venzar mixture on the respiration on *Pseudomonas aeruginosa* and *Arthrobacter* sp. strains

by Karpiak, Iwanowski [10], can stimulate or inhibit disproportionately oxygen and oxygen-free processes.

Generally, one can say that 50% of the bacteria examined were susceptible to the presence of venzar, which manifested itself by a disturbance in respiration processes. In this group were present forms of *Corynebacterium* sp., *Serratia* sp., eight strains of genus *Pseudomonas* sp. and two strains of genus *Bacillus* sp. The other bacterial strains of genus: *Bacillus*, *Pseudomonas* and *Arthrobacter* utilized venzar as

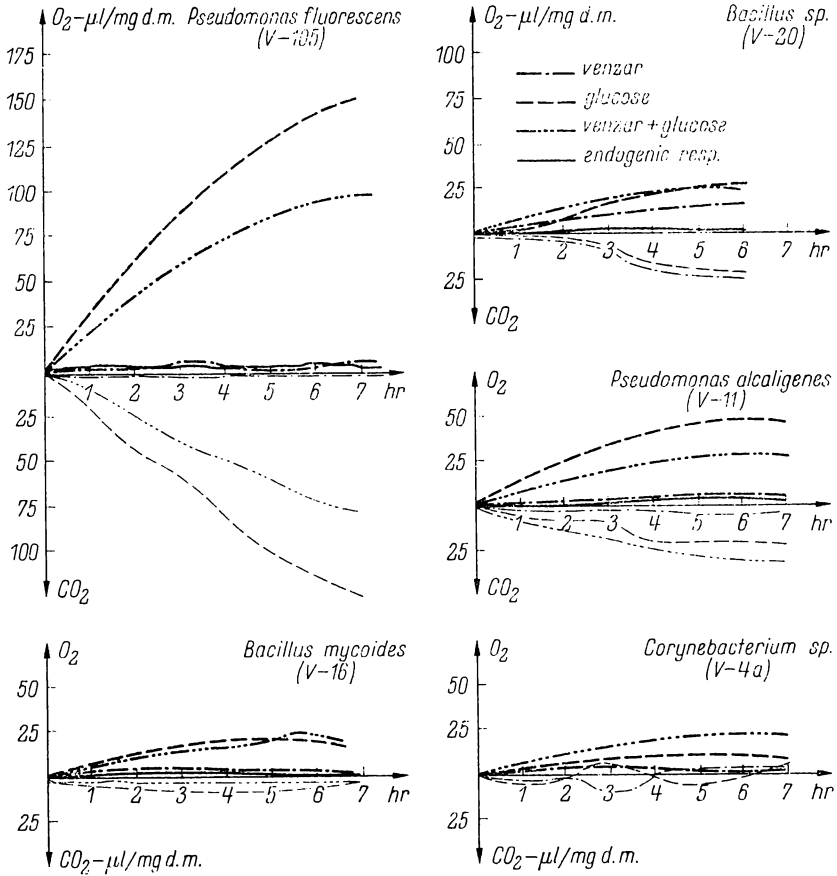


Fig. 5. The effect of glucose and a mixture of glucose and venzar on the respiration of various bacterial strains

a substrate in processes of oxygen respiration, thus contributing to its removal from the medium.

The obtained results stress the necessity of further investigations on microbial degradation of herbicides, which we will carry out in our successive experiments whose justification is also found in the studies by: Alexander [1, 2], Miyamoto and Ontavoi [13], Ballag [3, 4], Gołab, Althans [9], Sobieszcański [16].

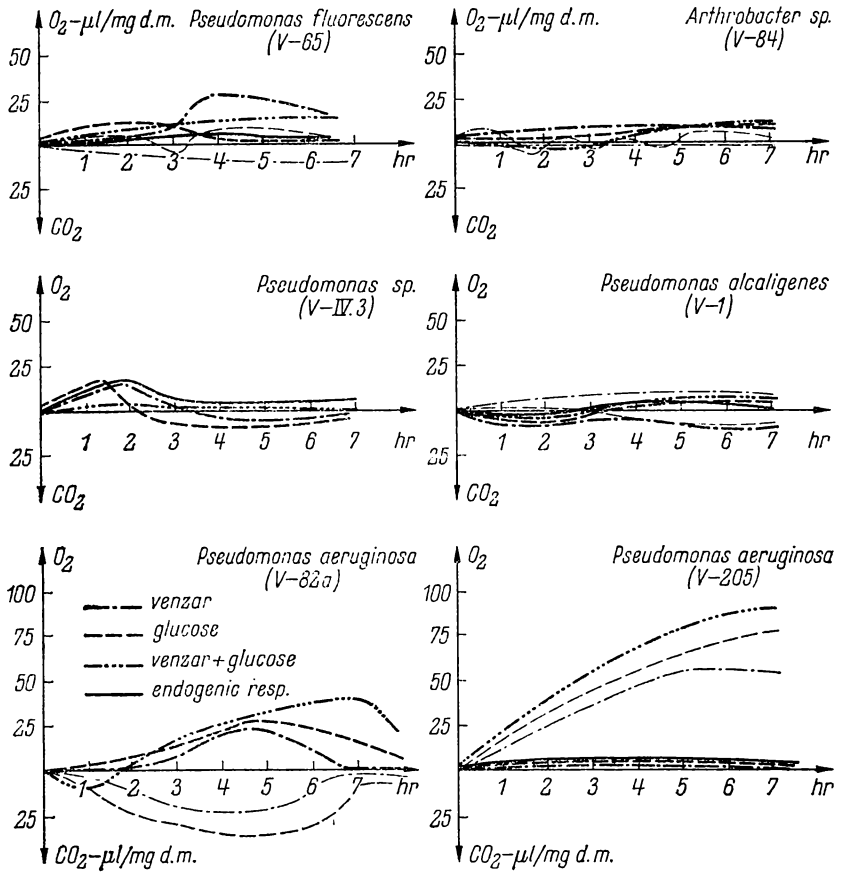


Fig. 6. The disturbance in respiration of *Pseudomonas* and *Arthrobacter sp.*

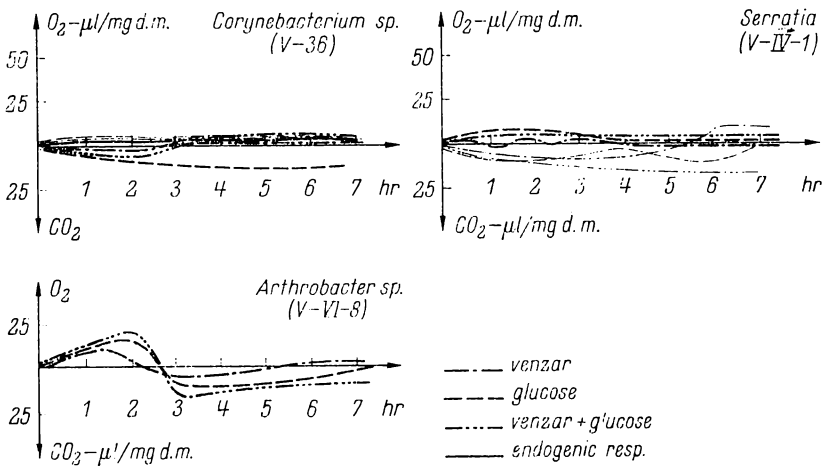


Fig. 7. The disturbance in respiration of *Corynebacterium sp.*

Table 2

The effect of venzar and glucose on gas exchange of the tested genera of bacteria and the decrement of venzar from the medium

Bacterial strains		RQ			Venzar residue	
		Respiration substrates				
N ^o	Genus	venzar 6 ppm	glucose 0.1%	% venzar + glucose	%	Rf of metabo - lites
V-217	<i>Bacillus</i> sp.	1.17	5.30	2.90	0	0.39 0.92
V-218	<i>Bacillus</i> sp.	0.91	1.40	1.09	70	_____
V-92	<i>Bacillus</i> sp.	0.98	1.27	1.05	20	_____
V-25	<i>Bacillus</i> sp.	1.12	1.27	0.42	25	_____
V-76	<i>Bacillus</i> sp.	1.20	1.12	1.40	20	_____
V-20	<i>Bacillus</i> sp.	1.27	0.81	-	0	_____
V-16	<i>Bacillus mycoides</i>	1.06	0.29	0.20	98	_____

V-80	<i>Pseudomonas fluorescens</i>	1.08	0.68	0.82	0	0.47 0.66 0.94
V-209	<i>Pseudomonas fluorescens</i>	0.70	irrational	-	0	0.25 0.40 _____
V-208	<i>Pseudomonas fluorescens</i>	0.64	0.47	2.10	20	0.29 _____
V-105	<i>Pseudomonas fluorescens</i>	1.30	0.67	0.75	100	_____
V-65	<i>Pseudomonas fluorescens</i>	0.68	0.84	-	95	_____
V-212	<i>Pseudomonas aeruginosa</i>	1.00	0.35	0.70	0	0.47 0.88 0.95
V-204	<i>Pseudomonas aeruginosa</i>	0.44	0.33	0.38	0	_____
V-211	<i>Pseudomonas aeruginosa</i>	1.30	1.00	-	0	0.17 0.37 _____
V-100	<i>Pseudomonas aeruginosa</i>	1.60	0.82	-	20	_____
VI-5	<i>Pseudomonas aeruginosa</i>	1.12	7.40	-	50	0.32 _____
V-206	<i>Pseudomonas aeruginosa</i>	0.91	0.52	-	50	0.32 _____
V-223	<i>Pseudomonas aeruginosa</i>	1.20	2.50	-	100	_____
V-82a	<i>Pseudomonas aeruginosa</i>	0.38	0.28	0.47	90	_____
V-205	<i>Pseudomonas aeruginosa</i>	1.40	4.40	-	0	_____
V-11	<i>Pseudomonas alcaligenes</i>	1.05	0.45	1.18	0	_____
V-1	<i>Pseudomonas alcaligenes</i>	1.17	1.00	-	0	_____
VIV-3	<i>Pseudomonas species</i>	-	-	-	0	_____

V-4a	<i>Corynebacterium</i> sp.	11.80	0.25	-	25	_____
V-84	<i>Corynebacterium</i> sp.	0.12	0.26	-	95	_____
V-36	<i>Corynebacterium</i> sp.	0.19	0.25	7.30	100	_____

V-216	<i>Arthrobacter</i> sp.	0.36	21.7	-	20	_____
VVI-8	<i>Arthrobacter</i> sp.	-	-	-	0	_____

V-5	<i>Micrococcus</i>	-	-	1.08	25	_____

VIV-1	<i>Serratia</i>	4.4	1.6	4.30	50	_____

V-10	<i>Nocardia</i>	2.07	1.12	-	20	_____

CONCLUSIONS

1. Bacteria of genera: *Bacillus*, *Pseudomonas*, *Nocardia*, *Corynebacterium*, *Serratia*, *Micrococcus* and *Arthrobacter* are susceptible to high doses of venzar (100 nad 1000 ppm) in the medium.

2. Most bacteria of genera: *Pseudomonas* *Bacillus* and *Arthrobacter* can utilize this herbicide as a carbon source in energetic processes, thus contributing to the removal of venzar from the medium.

3. Venzar at 6 ppm stimulated respiration processes in bacteria of genera: *Bacillus*, *Pseudomonas*, *Arthrobacter*.

4. In some of genera of bacteria (*Corynebacterium* sp., *Serratia* sp., *Nocardia* sp., *Micrococcus* sp.) venzar disturbed respiration processes.

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MIKROBIOLOGICZNA TRANSFORMACJA HERBICYDÓW W PROCESACH
KATABOLICZNYCH

CZEŚĆ I. WPŁYW VENZARU NA AKTYWNOŚĆ ODDECHOWĄ
WYBRANYCH RODZAJÓW BAKTERII GLEBOWYCH

Instytut Przechowalnictwa i Technologii Żywności AR
we Wrocławiu

Streszczenie

Badano 32 szczepy bakterii wyodrębnione z różnych gleb uprzednio perkolorowanych wodą zawierającą niską zawartość wenzaru [tab. 1]. Szczepy te należały do rodzajów: *Pseudomonas*, *Bacillus*, *Corynebacterium*, *Serratia*, *Nocardia*, *Micrococcus* i były odporne na dawkę 100 i 1000 ppm wenzaru w podłożu. Posługując się bezpośrednią metodą Warburga badano proces oddychania szczepów bakterii oznaczając ilość pobieranego tlenu i wydzielanego dwutlenku węgla w obecności wenzaru, glukozy i mieszaniny obu substancji jako substratów oddechowych. Wenzar okazał się dobrym substratem oddechowym dla większości bakterii z rodzajów: *Pseudomonas*, *Bacillus*, *Arthrobacter* i stymulował oddychanie tych szczepów bakterii. Pozostałe szczepy bakterii wykazały w obecności wenzaru niską aktywność oddechową i zaburzenia procesów wymiany gazowej.

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